Short-Term Effects of Oral Feeding Jujube Ziziphus Solution before a Single Session of Circuit Resistance Exercise on Apoptosis of Human Neutrophil

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ABSTRACT
The aim of present research was the effect of short-term use of edible solution of jujube one week before a single session of circuit resistance exercise on neutrophil apoptosis of male students of physical education. 14 young male volunteer students were divided into two groups of placebo (n=7, aged 24.50±2.50, height 171.17±1.70, and weight 67.51±4.92) and jujube solution (n=7, aged 25.25±1.31, height 179.75±3.63, weight 74.07±5.78) at random, performing one circuit resistance exercise (9 moves/stops, 30 seconds for each exercise, 3 nonstop sets with a 3-minute active recession between sets, 10-14 repetitions, and an intensity of 70% maximum repetition). Subjects received placebo and jujube solutions (0.5 g/kg body weight in 2.5 cc of distilled water as long as 7 days) at certain times and double-blind. Blood samples were collected 30 minutes before, immediately, and 2 hours after the exercise for separation and counting the number of neutrophils, and neutrophil apoptosis was determined through AnexinV-FITC kit and flow cytometric method. The results indicated the significant response of initial apoptosis neutrophils to one course of activity by jujube group. However, different responses were observed between two groups of placebo and jujube during the recession after training. Unlike placebo group, the response was significantly lower with a greater decrease after training in jujube group. However, necrosis/delayed apoptosis neutrophils significantly increased in placebo group after 2 hours, while this change was not observed in jujube group. At the end of a 2-hour recession, delayed apoptosis neutrophils significantly decreased in both placebo and jujube groups. The present findings indicate that one-session resistance activity is not very effective after pretreatment with jujube solution during one week. Perhaps this useful effect of jujube can be explained by existing glucose compounds and amino acids which could provide neutrophils with proper nutrition source in comparison with placebo. It is possible that jujube resulted in improvement of neutrophil’s antioxidant capacity, because it has proper antioxidant materials. Therefore, it might be stated that loading with edible jujube solution can be effective in inhibiting apoptosis at least for one week.

Key Words: Oral Jujube Solution, Neutrophil, Early Apoptotic, Late Apoptosis, Necrotic Neutrophils.

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INTRODUCTION
Neutrophils play a critical role in the first line of defense against pathogens. They rapidly migrate to the infection site, ingest the pathogens, release reactive oxygen species (ROS) to kill the pathogens, and even release their own DNA to form extracellular traps (1).

It is important that neutrophil activity be tightly regulated to prevent perpetuation of inflammation (2). Despite having a life span of 5 days under physiological conditions (3), neutrophil longevity is extended in inflammatory environments by delaying apoptosis/programmed cell death. Nevertheless, once an episode of acute neutrophilic inflammation is complete, it is essential that neutrophil recruitment be halted and that recruited neutrophils undergo apoptosis, before disposal of the apoptotic cells by surrounding phagocytes such as macrophages, to ensure efficient resolution of inflammation (4).

The natural process of neutrophil apoptosis resolution followed by phagocytosis is impaired in numerous human inflammatory disease states with delayed apoptosis (5-8) seen in exercise conditions (9). Recently, Mooren et al. (2012) reported that neutrophil apoptosis was delayed after both marathon run and intensive laboratory exercise tests evaluated with annexin V labeling and flow cytometry measuring. Apoptosis delay was accompanied under these exercise conditions by enhanced intracellular calcium transients and decreased glutathione levels. However, previous human studies show that repeated acute exercise increases apoptosis and reduces mitochondrial membrane potential (ΔΨm) in neutrophils (10-12).

Besides, a single bout of exercise induces an inflammatory response that is similar to that brought about by infection or trauma (13). A prolonged inflammatory state has detrimental health effects and predisposes to a number of chronic diseases and health conditions (13).

The control of granulocyte cell death and subsequent clearance of the apoptotic cell is not only crucial for maintaining homeostasis, but also paramount to an efficient resolution of inflammation (14). Neutrophils undergo spontaneous apoptosis at inflamed sites (15).

There are contradictory and limited reports about effect of glutamine supplementation in acute exercise and neutrophil response to it; it is reported in a review article that glutamine supplementation is beneficial for immunological function only in clinical situations and it wasn’t proven that it is effective for prevention of immune cells function following exercise (16, 17); so that, the effect of glutamine supplementation wasn’t significant during and after exercise (18). But some reports said that glutamine supplementation had an either significant effect on neutrophil function due to exercise (19) or a protective effect on neutrophil apoptosis due to acute exercise (20, 21), and also, an study without exercise intervention reported a protective effect against events with stimulation and execution of apoptosis on human and rat neutrophil (22).

Recently, herbal medicine and medical plants are widely used for the treatment of diseases and weight management (23, 24). Zizyphus/Jujube/Red Date/Annab is a plant of Khamancea family that contains different kinds of proteins, sugars, and also amino acids such as alanine, aspartic acid, glutamic acid (25), can participate in glutamine formation. Ghanbari-Niaki et al. (2013) investigated the effect of aerobic training, with or without Zizyphus Jujuba water extraction, on fundus nesfatin-1, ATP, HDL-c, and LDL-c concentrations (26) and liver and plasma Nesfatin-1 (27) in female rats and concluded that exercise and using jujube extraction may prevent over-weight and

cardiovascular diseases (26) and it could be considered as an anti-appetite herb (27).

It is believed the dried fruits of Zizyphus jujube are anodyne, anticancer, pectoral, refrigerant, sedative, stomachic, styptic and tonic and immune response enhancer (28-30). It has anti-inflammatory and anti-diabetic effects (26).

However, there are studies about beneficial effect of jujube extraction and some it’s compounds on immunological function (31-35) and also it is presented about protective effects of it on oxidant status (36-38), but there are a few studies about effects of jujube extraction on apoptosis induction in tumor cells (39, 40) and an report exist about the effects of jujube extraction on apoptosis status in heart muscles in response to acute exercise (41), but there isn’t any evidence about neutrophil specially.

So, the main question of this study is what the short-term effects of oral feeding Ziziphus Jujube solution is before a single session of circuit resistance exercise on apoptosis of human neutrophil cells measuring with Anexin V labeling and flow cytometry method.

**MATERIALS AND METHODS**

**Subjects.** The present study was approved by the Research Ethics Committee of the School of Medical Sciences of Tarbiat Modares University (Iran), and conducted in accordance with policy statement of the Declaration of Iranian Ministry of Health. Written informed consent was obtained from 14 young healthy male students. All subjects were asked to complete a medical examination as well as a medical questionnaire to ensure that they were not taking any regular medications for 1 month ago; no smoking, no alcohol, no regular exercise in the past 2 months, and free of cardiovascular or metabolic diseases, and also, recent symptoms of upper respiratory tract infection in 1 month before beginning the test. The volunteers were assigned randomly to 2 groups (n=7) including a Circuit Resistance Exercise group with placebo (CREP) (age: 24.5±2.5 years, height: 171.17±1.7 cm, weight: 67.51±4.92 kg) and Circuit Resistance Exercise group (n=7) with jujube solution (age: 25.25±1.31 years, height: 179.75±3.63 cm, weight: 74.04±5.78 kg) (CRES).

**Exercise Protocol and Blood Collection.** Participants were taken to the weight room three times before the main trial. Strength test was performed on the first and second visits to determine one repetition maximum (1-RM) of all the participants for each of the 9-resistance exercises, employed in the study. The 1-RM value was determined by trial and by adding or removing weights after each attempt as per required. The subjects were allowed to take as long time as they felt necessary to recover from each attempt. The subjects completed a practice session to ensure that each participant was able to complete the entire exercise session on the third visit, and also to confirm that the weight lifting was producing fatigue at the end of the session. This was confirmed by visual and verbal feedback from the participants (42). Records of 1-RM are presented in table 1.

Subjects in both groups arrived in the place of test at 08:00 and rested for about 30 minutes, all subjects performed the circuit resistance exercise in two circles at 08:30 at the same time. Each circle contained 9 exercises (crunch, back extension, biceps curl, triceps press, knee extension, knee curl, standing calf raise, chest press, seated row, all with machines). The test included three non-stop circuits with 3-minute active rests between circuits. Each exercise performed 30 s (about 10-14 repeats) with 75% one repeat maximum (1RM). The exercise protocol is drawn in figure 1.

Table 1. Exercise’s 1-RM Records of participants

<table>
<thead>
<tr>
<th>Variables (kg)</th>
<th>groups</th>
<th>Mean ± SE</th>
</tr>
</thead>
<tbody>
<tr>
<td>1 crunch</td>
<td>placebo</td>
<td>127.12 ± 8.66</td>
</tr>
<tr>
<td></td>
<td>Jujube solution</td>
<td>129.49 ± 10.21</td>
</tr>
<tr>
<td>2 back extension</td>
<td>placebo</td>
<td>199.76 ± 24.46</td>
</tr>
<tr>
<td></td>
<td>Jujube solution</td>
<td>270.77 ± 82.12</td>
</tr>
<tr>
<td>3 biceps curl</td>
<td>placebo</td>
<td>54.01 ± 5.38</td>
</tr>
<tr>
<td></td>
<td>Jujube solution</td>
<td>62.55 ± 4.49</td>
</tr>
<tr>
<td>4 triceps press</td>
<td>placebo</td>
<td>57.39 ± 4.29</td>
</tr>
<tr>
<td></td>
<td>Jujube solution</td>
<td>60.97 ± 4.67</td>
</tr>
<tr>
<td>5 knee extension</td>
<td>placebo</td>
<td>156.15 ± 13.07</td>
</tr>
<tr>
<td></td>
<td>Jujube solution</td>
<td>173.69 ± 11.27</td>
</tr>
<tr>
<td>6 knee curl</td>
<td>placebo</td>
<td>100.43 ± 9.67</td>
</tr>
<tr>
<td></td>
<td>Jujube solution</td>
<td>110.35 ± 18.12</td>
</tr>
<tr>
<td>7 standing calf</td>
<td>placebo</td>
<td>152.63 ± 15.20</td>
</tr>
<tr>
<td>raise</td>
<td>Jujube solution</td>
<td>166.08 ± 10.25</td>
</tr>
<tr>
<td>8 chest press</td>
<td>placebo</td>
<td>73.29 ± 3.38</td>
</tr>
<tr>
<td></td>
<td>Jujube solution</td>
<td>77.56 ± 9.83</td>
</tr>
<tr>
<td>9 seated row</td>
<td>placebo</td>
<td>126.46 ± 7.73</td>
</tr>
<tr>
<td></td>
<td>Jujube solution</td>
<td>149.15 ± 11.45</td>
</tr>
</tbody>
</table>

Figure 1. Exercise protocol

The semi-dried fruits of Zyzyphus Jujube were washed, and their seeds were separated and removed from soft red parts. The samples were dried in 50°C and were grounded into powder in a mortar. The groups received oral jujube solution (0.5 g/kg body weight in 2.5cc distilled water) and placebo (2.5cc/kg of body weight in distilled water sweetened by sugar without the calories and colored by food dyes) daily.
for one week in double blind manner without any physical training during this period. First, peripheral venous blood samples were drawn (8th day and after 12 hours of overnight fasting) at 08:30. Second, blood samples were taken immediately after exercise at 09:00, then subject sitted for 120 minutes. Third, blood samples were drawn at 11:00. The research design and blood collection is drawn in figure 2.

**Reagents.** Anticoagulant ACD [mixture of citric acid (Sigma Aldrich, Germany), sodium citrate (Sigma Aldrich, Germany), dextrose (Sigma Aldrich, Germany), and Deionized Sterile Distilled Water/dsdH2O (Baharafshan, B.I.R.D, Iran)], Dextran 6% [combination of dextran with at least 100000 MW (grade B, BDH lab, GPR™, England), 0.9% NaCl (Sigma Aldrich, Germany), and dsdH2O], 0.6 M KCl [mixture of KCl (Sigma Aldrich, Germany), and dsdH2O], Phosphate buffered saline/PBS solution [combination of NaCl, KCl, Na2HPO4 (Sigma Aldrich, Germany), KH2PO4 (Sigma Aldrich, Germany), and dsdH2O] with pH 7.4 and Autoclaved, Hank's Balanced Salt Solution/HBSS without Ca & Mg [NaCl, KCl, Na2HPO4, KH2PO4, NaHCO3 (Sigma Aldrich, Germany), glucose (Sigma Aldrich, Germany), and dsdH2O] with pH 7.4 and Autoclaved, Ficoll-Hyphaque 10.77 (Baharafshan, B.I.R.D, Iran).

**Neutrophil isolation.** Neutrophils were purified from venous blood treated with ACD from healthy volunteers by 3-steps: Dextran sedimentation, Hypotonic lysis, and Ficol sedimentation (43). Briefly, solution of 6% Dextran & 0.9% NaCl added into the mixture of ACD & blood and after adequate mixing, it stood down at room temperature until separation was complete. Then, the yellowish supernatant was separated and centrifuged using a low brake. After breaking the pellet with discarding the supernatant and re-suspension in dsdH2O at 20 s, it mixed with 0.6M KCl and the solution diluted with PBS. Then, it centrifuged using a high brake. The supernatant discarded and the pellet re-suspended in PBS. The cell suspension layered over Ficoll-Hyphaque and centrifuged using a low brake. Final, the supernatant discarded and the neutrophils pellet resuspended in HBSS. Trypan Blue Viability Test using hemocytometer chamber was performed to assay cell viability and it was ≥ 96%.

**Apoptosis assessment.** Apoptotic cell death was assessed using FITC-conjugated Annexin-V/PI assay kit (BioVision, USA) by flow cytometry (COULTER, EPICS XL-MCL, by Beckman Coulter, USA). Briefly, after collection of 5 x 10⁵ cells by centrifugation, it re-suspended in 500 μl of 1X Binding Buffer; then, 5 μl of Annexin V-FITC and PI added to suspension; and it

incubated at room temperature for 5 min in the dark. Finally, Annexin V-FITC binding analyzed by flow cytometry (Ex = 488 nm; Em = 530 nm) using FITC signal detector (usually FL1) and PI staining by the phycoerythrin emission signal detector (usually FL2). The percentage of cells stained with Annexin-V only was estimated as early apoptosis; the percentage of cells stained with both Annexin-V and propidium iodide was estimated as late apoptosis or necrotic phase.

**Statistical analysis.** Repeated measure (two-way) ANOVA was used to determine the effects of TIME and SOLUTION by SPSS software at a significance level of p = 0.05. All data were presented as means with standard error of mean.

**RESULTS**

**Neutrophil Counts.** Mauchly's test of sphericity for Neutrophil Counts is met (Mauchly's W = 0.99, p = 0.97). The main effect of TIME with assumption of sphericity was significant (F = 45.405, p = 0.001) and this effect was linear (F = 84.9, p = 0.001). But, the main effect of SOLUTION wasn’t significant (F = 0.018, p = 0.9); besides, the interaction of TIME and SOLUTION with assumption of sphericity was significant (F = 3.56, p = 0.01) and this effect was quadratic (F = 10.015, p = 0.008) [Graph 1].

**Alive Neutrophils:** Mauchly's test of sphericity for Alive Cells is met (Mauchly's W = 0.67, p = 0.109). The main effect of TIME with assumption of sphericity was significant (F = 11.92, p = 0.001) and this effect was quadratic (F = 15.8, p = 0.002). But, the main effect of SOLUTION wasn’t significant (F = 0.01, p = 0.929); besides, the interaction of TIME and SOLUTION with assumption of sphericity was significant (F = 3.56, p = 0.048) and this effect was linear (F = 4.95, p = 0.049) [Graph 2].
**Graph 2.** Alive Cell in Response to Short-term Effect of Solution Feeding.

**Early Apoptotic Neutrophils:** Mauchly's test of sphericity for Early Apoptotic Cells was met (Mauchly's W = 0.89, p = 0.52). The main effect of TIME with assumption of sphericity was significant (F = 24.72, p = 0.001) and this effect was quadratic (F = 64.1, p = 0.001). Also, the main effect of SOLUTION was significant (F = 4.81, p = 0.049); besides, the interaction of TIME and SOLUTION with assumption of sphericity was significant (F = 3.91, p = 0.049) and this effect was linear (F = 5.002, p = 0.045) [Graph 3].

**Graph 3.** Early Apoptotic Cells in Response to Short-term Effect of Solution Feeding.

**Late Apoptotic/Necrotic Neutrophils:** Mauchly's test of sphericity for Late Apoptotic Cells is met (Mauchly's W = 0.64, p = 0.08). The main effect of TIME with
assumption of sphericity was significant ($F = 9.29, p = 0.001$) and this effect was quadratic ($F = 11.82, p = 0.005$). But, the main effect of SOLUTION wasn’t significant ($F = 0.4, p = 0.54$); besides, the interaction of TIME and SOLUTION with assumption of sphericity was significant ($F = 3.55, p = 0.048$) and this effect was linear ($F = 5.07, p = 0.049$) [Graph 4].

**DISCUSSION**

We saw that Neutrophil Counts increased significantly in response to circuit resistance exercise in both groups of Jujube Solution and Placebo. Many studies have shown that brief exercise increases the circulating neutrophil count (44, 45). Ghanbari-Niaaki and Tayebi (2013) reported that circulating neutrophil counts of male college students didn’t change significantly in response to a single circuit resistance exercise with 35% 1RM (46); but, Tayebi *et al.* (unpublished data) showed that a single circuit resistance exercise with 60% 1RM resulted a significant elevation in circulating neutrophil counts of male college students but not in young male weightlifters.

During very high intensity exercise lasting only 60 seconds the circulating granulocyte count increases and peaks 15 minutes post-exercise (47). It has also been highlighted that for brief exercise the granulocytosis is dependent on intensity (48). The circulating neutrophil count has been reported to increase by up to about 90% (49) after brief exhaustive exercise. It appears that the demargination of neutrophils by adrenaline is selective, because adrenaline infusion produces a neutrophilia that has a slightly higher percentage of segmented (mature) neutrophils (50), as we saw that Alive Cells increased significantly in placebo group.

Besides, the results of this study showed that Neutrophil Counts of placebo group decreased significantly 120 min after exercise rather than immediately after exercise and also was higher than pre-test. Some studies have demonstrated that only the number of lymphocytes and not neutrophils declined below the corresponding pre-exercise value 2 hours after physical activity (51), as we saw (under review data) an elevation in neutrophil counts after 2 hours of recovery from 1 bout eccentric exercise (only extension from curl up, 80% 1RM, 8-10 repeats, 6 sets). Recent studies suggest that the decline in leukocyte

![Graph 4. Late Apoptotic Neutrophils (%) in response to acute effect of solution feeding.](image-url)
number after exercise occurs due to the induction of leukocyte death (11). Moreover, the increase in DNA fragmentation depends on the intensity and duration of the exercise (19, 52). Exhaustive exercise increases DNA fragmentation in the lymphocytes and neutrophils of athletes (53-56). The study of Lagranha et al. (2004) showed an increase in the apoptosis of neutrophils obtained from immature and mature rats after a single session of exercise (21). Similar results were found by Levada-Pires et al. (2008) in neutrophils from elite athletes after competing in a triathlon (54).

Exercise mobilizes peripheral immune cells and the magnitude of this effect reflects the intensity and duration of the effort (57, 58). Intense training increases the susceptibility to infections (59) and this may result from impairment in neutrophil function or acceleration in the process of neutrophil death (11). Short-term exhaustive exercise or intensive treadmill exercise is known to induce DNA damage in lymphocytes in untrained individuals (60). Recently, Lagranha et al. (2004 and 2005) have demonstrated that a single session of exercise induces DNA fragmentation and mitochondrial membrane depolarization, increases expression of pro-apoptotic genes (bax and bcl-xS) and decreases expression of anti-apoptotic genes (bcl-xL) in rat neutrophils. (19, 21)

Apoptosis is a very complex phenomenon involving a regulated series of events, in part controlled by extracellular stimuli including cytokines and perhaps nutrient availability (61).

However the role of glucose and glutamine for apoptosis process remains poorly understood; but some studies mentioned that neutrophils utilize these metabolites at high rates (62).

Previous studies have revealed that jujube contains various constituents, including triterpenic acids, flavonoids, cerebrosides, amino acids, phenolic acids, mineral constituents, and polysaccharides and phytochemical studies of jujube fruits have shed some light on their biological effects, such as the anticancer, anti-inflammatory, anti-obesity, immune-stimulating, antioxidant, hepato-protective, and gastrointestinal protective activities and inhibition of foam cell formation in macrophages (63).

Our results showed that since neutrophil counts of jujube solution group increased during exercise, in placebo group didn’t have any change. But neutrophil counts of placebo group elevated during recovery as it suppressed in jujube solution group (interaction effect of time and solution). Besides, since living neutrophils of placebo group decreased significantly during exercise, it decreased insignificantly in jujube solution group, in other words, it suppressed. But living neutrophils of placebo group increased to baseline during recovery, so that of jujube solution group increased above baseline. On the other side, early apoptotic neutrophils of jujube solution group had a significant increase less than significant increase of placebo group, and also during recovery, it decreased to under baseline in jujube solution group, but it didn’t change in placebo group. Also, since late apoptotic neutrophils of jujube solution group suppressed during exercise and elevated in placebo group more than it. Although both groups had a significant decrease to baseline during recovery, it went under baseline in jujube solution group.

It is not clear that pretreated with oral Ziziphus Jujube solution in our research condition suppressed neutrophil apoptosis by what mechanism. Nonetheless, with respect to medicinal and effective composition container in jujube zizifus, it can be mentioned to some likely mechanisms of immune effect of jujube zizifus. Current researches about Ziziphus Jujube fruit show that its extraction have cytotoxic activity and can induce apoptosis in cancer cells and...
some tumor cells (40). They suggested that infusion of Ziziphus Jujube extraction to cultured cells caused shrinkage, and detachment of adhesive cells, HeLa and HEp-2. These effects increased by adding to extraction levels. Besides, it is noticed the anti-proliferative and tumor cell growth inhibitory effect of triterpene acids fraction container Ziziphus Jujube. It is maybe because of pretreated with Ziziphus Jujube solution could be prevented from that activity of neutrophils against physical stress. Huang et al. (2007) evaluated the effect of a Chinese mixture medicine container Ziziphus Jujube as immune stimulus of mice. The results showed that the mixture could prevent from decrease of white blood cell count caused by cyclophosphamide (cyclophosphamide-induced leucopenia) (64). Shen et al. (2009) observed that treating of mice with ethanol extraction of Ziziphus Jujube in two dosages of 100 and 200 mL/kg of body weight could prevent from decreasing activity of antioxidant levels (glutathione and superoxide dismutase) caused by carbon tetrachloride (ccl4-induced) (37). Ganachari, Kumar, and Bhat (2004) studied the effect of Ziziphus jujuba leaves extract on phagocytosis by human neutrophils and saw that the extract elevated motion of neutrophils from up section to under of filter as dose-related and decreased neutrophilic phagocytosis gently with increase of extract level (65). Yu et al. (2012) and Goyal et al. (2011) investigated the anti-inflammatory effects of Ziziphus jujube; and demonstrated that the triterpene acids fraction was the most active part of jujube through the inhibitory effects on the inflammatory cells activated by Euphorbia kansui and prostratin, a phorbol ester isolated from Euphorbia fischeriana (66); or suggested that it plays a protective role against experimental acute and chronic inflammatory reactions in rat, possibly by attenuating NOS activity (67). Also, Li et al. (2011) and Zhao et al. (2006) studied the immune stimulus effects of Ziziphus jujube; and reported that its polysaccharides fractions (ZSP) such as crude ZSP dramatically increased thymus and spleen indices in mice and enhanced the proliferation of splenocytes and peritoneal macrophages (68); and/or its pectic polysaccharides (Ju-B-2) had a significant activity in enhancing the effect of spleen cells proliferation at a higher dose (>30 μg/mL) (69). But, about its immunological effect in exercise and training doesn’t exist in any evidence. Only, Ghanbari-Niaki et al. (2013) studied the effect of aerobic training with and without Zizyphus Jujuba water extraction on liver and plasma Nesfatin-1 (27) and fundus Nesfatin-1, ATP, HDL-C, and LDL-C concentrations (26) of female rats and reported that exercise training with Zizyphus Jujuba water extraction probably prevent from overweighting and cardiovascular disease (26) and it could be considered as an anti-appetite herb (27). Tayebi et al. (unpublished data) investigated acute effects of oral feeding Jujube Ziziphus solution an hour before a single session of circuit resistance exercise (exactly such as this study protocol) on apoptosis of human neutrophil and observed that neutrophil apoptosis suppressed in Jujube Ziziphus solution.

According to nutrition sciences, jujuba consists of different nutrient and materials including sugars (Starch 21.8%, fructose 16%, glucose 9.6%, and sucrose 21.8%), Fat (19%), various amino acids (Glycine, histidine, leucine, iso-leucine, phenylalanine, proline, serine, threonine and etc.), glutamic acid, protein (4.5-5.6%), various minerals (Iron, sodium, potassium, zinc, manganese, sulfur and etc.) and vitamins (C, B1, B). Generally, jujube fruit is high in carbohydrates, especially fructose and glucose, which account for about 77% of its weight. Vitamins C, B complex, and A, as well as calcium, potassium, and other

mineral elements, have also been identified in jujuba fruit (70, 71).

As it was mentioned in the previous studies, glucose and glutamine are two of the most important fuel for the cell of immune system. The functionality of glutamine in rats’ neutrophils was particularly studied, and it was proven for the first time that the extent of glutamine consumption in these cells was higher than glucose (62). It seems that glutamine consumption has increased in apoptosis neutrophils. On the other hand, it has been indicated that glutamine postponed the occurrence of spontaneous apoptosis in neutrophils, and plasma concentration of glutamine was always positively correlated to MTP (22). A decline in MTP was detected as the necessary commitment of cells to apoptosis (55). Also, glutamine plasma was inversely externalized with annexin V binding to phosphatidylserine, and it was correlated with chromatin concentration of neutrophils in both rats and humans (22).

On the other hand, amino acids such as alanin, aspartic acid, especially glutamic acid can participate in synthesis of glutamine, and it seems that availability of these amino acids in body by its loading a week before exercise can be effective in glutamine synthesis. So it can be concluded that apoptosis suppression in Jujube Ziziphus group is because of glucose and glutamine availability resulted from loading of amino acids during a week before examination.

CONCLUSION
In conclusion, it is considered that this study examine the effects of supplementation with water solution of Jujube Ziziphus on apoptotic situation caused by exercise and antiapoptotic effect of it with resistance exercise for the first time. The results suggest that oral Jujube Ziziphus solution can be a proper supplement before resistance activities specially circuit resistance exercise because of its nutritional characteristics and values, because it can prevent from elevation of neutrophil apoptosis caused by circuit resistance exercise as it is related to occurrence of some intracellular events. Therefore, it is needed to conduct more researches to clarify proper mechanisms of inhibitory and anti-apoptotic effects of Jujube Ziziphus.

REFERENCES


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چکیده

هدف از تحقیق حاصل اثر مصرف کوتاه‌مدت محلول خوراکی عناب یک هفته قبل از یک هفته تمرین در افراد در دانشگاه تربیت مدرس تهران بود. در این تحقیق، 24 دانشجو داوطلب پسر جوان داوطلب بطور تصادفی به دو گروه تمرین مقاومتی دایره‌ای دارونما (1 نفر با سن 27/16 ± 2/74 و وزن 175/87 ± 2/74 و عدد (7) فرد) و به عنوان (2 نفر با سن 27/16 ± 2/74 و وزن 175/87 ± 2/74) که هر یک هفته دو گروه م臑یت و تمرین مالک آورده شدند. افراد در ساعت مشخص محلول‌های دارونما و عناب (5/0) گرم به آرزه کیلوگرم وزن بین در 7/5 مسی ای بین و هم‌اکنون 7/5‌وزن و 7/5ره صورت دو دوره در هفته کردند. نمونه‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انزیم‌های فیزیولوژی و انз...
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